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=> d his

     (FILE 'HOME' ENTERED AT 09:10:03 ON 22 SEP 2004)

     FILE 'MEDLINE, CAPLUS' ENTERED AT 09:11:00 ON 22 SEP 2004

     FILE 'MEDLINE, BIOSIS, CAPLUS' ENTERED AT 09:11:11 ON 22 SEP 2004
L1            18 S (RAPID?) (9A) (AMPLIF?) (9A) BIOPS?
L2             8 DUP REM L1 (10 DUPLICATES REMOVED)
L3          1642 S RAPID? AND (AMPLIF? OR PCR OR POLYMERASE (W)CHAIN) AND BIOPS?
L4          1642 S RAPID? AND (AMPLIF? OR PCR OR POLYMERASE (W)CHAIN) AND BIOPS?
L5           271 S L4 AND PY<1994
L6           138 DUP REM L5 (133 DUPLICATES REMOVED)

     FILE 'STNGUIDE' ENTERED AT 09:21:31 ON 22 SEP 2004

     FILE 'MEDLINE, BIOSIS, CAPLUS' ENTERED AT 09:22:36 ON 22 SEP 2004

     FILE 'STNGUIDE' ENTERED AT 09:22:36 ON 22 SEP 2004

     FILE 'MEDLINE, BIOSIS, CAPLUS' ENTERED AT 09:23:25 ON 22 SEP 2004
L7           325 S PHENOL (9A) (PCR OR POLYMERASE (W)CHAIN)
L8            45 S L7 AND PY<1994
L9            27 DUP REM L8 (18 DUPLICATES REMOVED)

     FILE 'STNGUIDE' ENTERED AT 09:28:18 ON 22 SEP 2004

     FILE 'MEDLINE, BIOSIS, CAPLUS' ENTERED AT 09:32:05 ON 22 SEP 2004
L10           53 S "WITHOUT" (9A) (ETHANOL OR ETOH) (9A) (PRECIP? OR PPT)
L11           37 DUP REM L10 (16 DUPLICATES REMOVED)

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L9 ANSWER 23 OF 27 CAPLUS COPYRIGHT 2004 ACS on STN
AN 1991:1750 CAPLUS
DN 114:1750
TI DNA isolated from plastic embedded tissue is suitable for PCR
AU Gruenewald, K.; Feichtinger, H.; Weyrer, K.; Dietze, O.; Lyons, J.
CS Dep. Intern. Med., Univ. Innsbruck, Austria
SO Nucleic Acids Research (**1990**), 18(20), 6151
CODEN: NARHAD; ISSN: 0305-1048
DT Journal
LA English
AB Plastic embedding of routinely formalin fixed tissues preserves excellent morphol. and is used in many centers especially for routine histol. of bone marrow biopsies. Since recent reports have shown the feasibility of polymerase chain reaction (PCR) amplification of DNA isolated from fixed paraffin embedded tissue, DNA was extracted from bone marrow trephine biopsies fixed in 10% formalin, decalcified in EDTA, embedded in Technovit 700 (hydroxyethylmethacrylate; HEMA; Kulzer, FRG) and stored between 2 mo and 4 yr at room temperature to test also for the influence of different ds.p. of the embedding medium. This report describes 2 alternate methods used to extract the DNA from plastic embedded tissue for subsequent amplification by PCR. One method uses proteinase K digestion and extns. with **phenol**/chloroform/isoamyl alc. prior to **PCR** while the 2nd involves direct amplification of proteinase K digested supernatants. Length of proteinase K digestion and temperature were found to influence DNA recovery dramatically. Different storage times i.e. different ds.p. of the embedding medium HEMA, had no influence on DNA yield or degradation DNA isolated by either method 1 or 2 was suitable for amplification by PCR. The second method was used to amplify a 526 bp fragment of a Gs protein α chain gene.

L9 ANSWER 9 OF 27 MEDLINE on STN DUPLICATE 6
AN 93216961 MEDLINE
DN PubMed ID: 8385159
TI Simple technique for detecting RNA viruses by PCR in single sections of wax embedded tissue.
AU Woodall C J; Watt N J; Clements G B
CS Regional Virus Laboratory, Ruchill Hospital, Glasgow.
SO Journal of clinical pathology, **(1993 Mar)** 46 (3) 276-7.
Journal code: 0376601. ISSN: 0021-9746.
CY ENGLAND: United Kingdom
DT Journal; Article; (JOURNAL ARTICLE)
LA English
FS Abridged Index Medicus Journals; Priority Journals
EM 199305
ED Entered STN: 19930521
Last Updated on STN: 19970203
Entered Medline: 19930506
AB The detection of specific RNA species in wax-embedded tissue sections using the polymerase chain reaction (PCR) means that gene expression can be studied and RNA viruses detected in stored histological tissue samples. This technique potentially allows the distribution of gene expression and viral replication to be studied in finely subdivided tissues. A technique is presented that has been used successfully to detect short RNA target sequences (130-420 bases) from proto-oncogene Abelson, human enteroviruses, and the sheep retrovirus Maedi-Visna virus using RNA PCR in single wax sections (20-30 microns). Various tissues were used which had not been deliberately prepared for this purpose. In a simple procedure hot xylene dewaxing is followed by acid **phenol** extraction of RNA and RNA **PCR**.

L6 ANSWER 22 OF 138 MEDLINE on STN DUPLICATE 15
AN 93358930 MEDLINE
DN PubMed ID: 8354305
TI Detection of Helicobacter pylori in gastric **biopsy** tissue by **polymerase chain** reaction.
AU Wang J T; Lin J T; Sheu J C; Yang J C; Chen D S; Wang T H
CS Department of Internal Medicine, National Taiwan University Hospital, Taipei.
SO European journal of clinical microbiology & infectious diseases : official publication of the European Society of Clinical Microbiology, **(1993 May)** 12 (5) 367-71.
Journal code: 8804297. ISSN: 0934-9723.
CY GERMANY: Germany, Federal Republic of
DT Journal; Article; (JOURNAL ARTICLE)
LA English
FS Priority Journals
EM 199309
ED Entered STN: 19931008
Last Updated on STN: 19931008
Entered Medline: 19930921
AB To evaluate the sensitivity of a **polymerase chain** reaction (**PCR**) assay using nested primers in detecting Helicobacter pylori, gastric tissue **biopsy** specimens were collected on endoscopy from 17 patients with a duodenal ulcer. DNA was extracted by phenol/chloroform treatment or boiling in water, and then subjected to a nested **PCR** using two primer pairs from the urease gene of Helicobacter pylori. Fourteen of the 17 patients were positive for Helicobacter pylori using DNA samples extracted by either method. The **PCR** results correlated well with the results of an enzyme immunoassay to detect IgG antibody. However, there were two culture negative patients. The three **PCR** negative patients were both culture negative and serologically negative. DNA from 9 of the 14 patients was randomly selected and subjected to semiquantification by serial dilutions, and then **PCR**. The results showed that phenol/chloroform extraction yielded 10-1000 times more DNA than the boiling method. It is concluded that the **PCR** assay is a **rapid** and sensitive method for detecting Helicobacter pylori, and that phenol/chloroform extraction is superior to simple boiling in obtaining DNA samples for **PCR**.

L6 ANSWER 70 OF 138 MEDLINE on STN DUPLICATE 48
AN 92202313 MEDLINE
DN PubMed ID: 1313040
TI A simple and **rapid** method of high quantity DNA isolation from cervical scrapes for detection of human papillomavirus infection.
AU Gopalkrishna V; Francis A; Sharma J K; Das B C
CS Division of Molecular Oncology, Institute of Cytology and Preventive Oncology (ICMR), New Delhi, India.
SO Journal of virological methods, **(1992 Jan)** 36 (1) 63-72.
Journal code: 8005839. ISSN: 0166-0934.
CY Netherlands
DT Journal; Article; (JOURNAL ARTICLE)
LA English
FS Priority Journals
EM 199204
ED Entered STN: 19920509
Last Updated on STN: 19920509
Entered Medline: 19920428
AB Infection with human papillomavirus (HPV) is an important etiological factor in the development of cervical cancer, and detection of the viral genome is of prognostic importance, particularly for preneoplastic lesions. We developed a simple, easy and efficient non-organic method of DNA extraction from cervical scrapes for reliable detection of HPV DNA sequences. The method involves incubation of cell nuclei in higher concentration of proteinase K at 65 degrees C for 2.5 h. Following prolonged incubation at higher temperature, the enzyme is autoinactivated and the DNA isolated can be used directly for analysis without further purification. The recovery of DNA is more than 95% and it can be easily cleaved by restriction enzymes and is suitable for **amplification** by the **polymerase chain** reaction (**PCR**). The whole procedure is carried out in a single Eppendorf tube and a large number of specimens can be processed at a time without any error of handling. DNA extracted from a single smear sample is sufficient to conduct as many as four different molecular biology tests. This provides an opportunity for verification of sensitivity, specificity and reliability of each test for diagnosis of HPV infection without resorting to **biopsy**.

L6 ANSWER 124 OF 138 MEDLINE on STN DUPLICATE 81
AN 89356258 MEDLINE
DN PubMed ID: 2548820
TI A **rapid** procedure to identify newborn transgenic mice.
AU Lin C S; Magnuson T; Samols D
CS Department of Genetics, School of Medicine, Case Western Reserve University, Cleveland, OH 44106.
NC AR20618 (NIAMS)
P30CA43703 (NCI)
SO DNA (Mary Ann Liebert, Inc.), **(1989 May)** 8 (4) 297-9.
Journal code: 8302432. ISSN: 0198-0238.
CY United States
DT Journal; Article; (JOURNAL ARTICLE)
LA English
FS Priority Journals
EM 198910
ED Entered STN: 19900309
Last Updated on STN: 19980206
Entered Medline: 19891006
AB We have developed a **rapid** procedure to identify newborn transgenic mice containing foreign genetic material in their genome. The protocol involves collagenase digestion of a small amount of tail tissue which can be taken very early after birth, phenol and chloroform extraction, **polymerase chain** reaction, and polyacrylamide gel electrophoresis. The entire procedure, from tissue **biopsy** to final results, can be completed in 1 day.

ANSWER 119 OF 138 MEDLINE on STN DUPLICATE 79

AN 90104461 MEDLINE

DN PubMed ID: 2557850

TI Method of extracting DNA from fine needle aspirates of human solid tumors for Southern blot analysis.

AU Harnett P R; Greenberg M L; Tattersall M H; Kefford R F

CS Medical Oncology Unit, Institute of Clinical Pathology, Westmead, New South Wales, Australia.

SO Analytical and quantitative cytology and histology / the International Academy of Cytology [and] American Society of Cytology, **(1989 Dec)** 11 (6) 375-8.
Journal code: 8506819. ISSN: 0884-6812.

CY United States

DT Journal; Article; (JOURNAL ARTICLE)

LA English

FS Priority Journals

EM 199002

ED Entered STN: 19900328
Last Updated on STN: 19900328
Entered Medline: 19900222

AB A **rapid**, simple, convenient method for extracting DNA from fine needle aspiration (FNA) samples of human solid tumors for Southern blot hybridization studies is described. After the preparation of an air-dried cytologic smear, the remaining sample in the needle was rinsed directly into a test tube for DNA extraction. The extraction procedure, in which manipulation of the sample is minimized, produced sufficient DNA for Southern blot analysis within 24 hours of the FNA **biopsy** in the ten consecutive cases studied. The DNA bound to the nylon membranes can be washed and reexamined with a variety of probes, allowing studies of lymphoid cell lineage, oncogene **amplification** or tumor progression. The assessment of cellularity on the cytologic specimen at the time of FNA provided a reliable guide to the need for further passes to obtain sufficient cells for DNA hybridization; the cytologic diagnosis could also be made on the smears.

ANSWER 14 OF 27 MEDLINE on STN

AN 93187424 MEDLINE

DN PubMed ID: 1293226

TI Comparison of polymerase chain reaction and IDEIA Chlamydia in detection of Chlamydia trachomatis from first-voided urine of male urethritis patients.

AU Komeda H; Deguchi T; Tada K; Yamamoto H; Kanematsu E; Iwata H; Okano M; Ito Y; Saito A; Ban Y; +

CS Department of Urology, Gifu University School of Medicine.

SO Kansenshogaku zasshi. Journal of the Japanese Association for Infectious Diseases, **(1992 Oct)** 66 (10) 1473-8.
Journal code: 0236671. ISSN: 0387-5911.

CY Japan

DT Journal; Article; (JOURNAL ARTICLE)

LA Japanese

FS Priority Journals

EM 199304

ED Entered STN: 19930416
Last Updated on STN: 19930416
Entered Medline: 19930402

AB We have reported a method for detection of Chlamydia trachomatis by polymerase chain reaction (PCR) with two oligonucleotides based on sequences within the major outer membrane protein gene from C. trachomatis serovar L2. In the previous report, in addition to treatment of the mixture of first-voided urine (FVU) sediment and 1 ml of urine with proteinase K. DNA purification by **phenol** extraction was necessary for preparation of template DNA for **PCR**. In this study, FVU sediment was suspended in 1 ml of Chlamydiazyme dilution buffer and a part of the suspension was treated with proteinase K for DNA extraction. The DNA extraction solution could be used as template for **PCR** without purification of DNA by **phenol** extraction.
One hundred FVU specimens obtained from male urethritis patients were examined with the two methods (PCR and IDEIA) for detection of C. trachomatis. In 33 of 100 specimens, the DNA fragments of C. trachomatis was amplified by the PCR and in 32 of 100, the chlamydial antigen was detected by IDEIA. The positive and negative coincidence rate of the PCR to IDEIA were 93.8% (30.32) and 95.6% (65/68) respectively, resulting in a high overall coincidence rate at 95%. Thus, the improved method with PCR using FVU as a specimen is proved to be a useful, non-invasive diagnostic tool for diagnosis of chlamydial urethritis.